

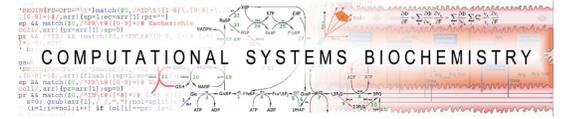
# Are enzyme costs minimized in evolution?

Enzyme size, efficiency and turnover as a mean for better flux predictions in FBA



Andreas Hoppe<sup>1</sup>, Christine Richter<sup>1</sup>, Hermann-Georg Holzhütter<sup>1</sup>

<sup>1</sup>Computational Systems Biochemistry, Institute of Biochemistry, Charité, Universitätsmedizin Berlin



**Summary.** Presumably, the costs of enzymes are minimized in the course of evolution, e.g. enzymes are as small (to minimize the effort for protein synthesis), as efficient (to minimize the necessary amount of enzymes for the required catalyzed flux), and with as low a degradation rate (to minimize the need to replenish the enzyme) as possible. However, as other factors (e.g. effective regulation

mechanisms) are critical for the cell's fitness it remains to be proved that costs have indeed an impact on the prevalence of enzymes throughout evolution. Here, the sequence lengths of the enzymes for central carbohydrate pathways is compared for selected classes in the tree of life in order to investigate the evolutionary preference of small enzymes. To assess the role of the different

cost components the effect on the cost estimates is analyzed for two alternative pathways which yield glyceraldehyde phosphate and energy equivalents from activated glucose. As a concluding prospect, a flux-balance optimization procedure based on the minimization of required enzyme costs is presented.

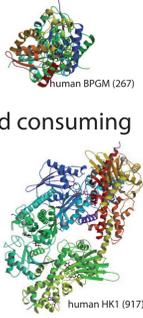
## Sequence length (by active site)

- fits perfectly in cost concept: **Investment**  
 - protein synthesis is major cell's energy consumer  
 - evolutionary pressure on protein size

### Constraints:

- Stable folding
- Protective sequences (Protrypsin)
- Coupling of free energy producing and consuming chemical reaction
- Membrane position
- Allosteric mechanisms
- Stable binding of prosthetic groups
- Regulatory sites
- Reaction chamber (proteasome)

**Data availability:** perfect (uniprot+PDB)



## Efficiency (turnover number)

fits relatively well in cost concept: **Productivity**

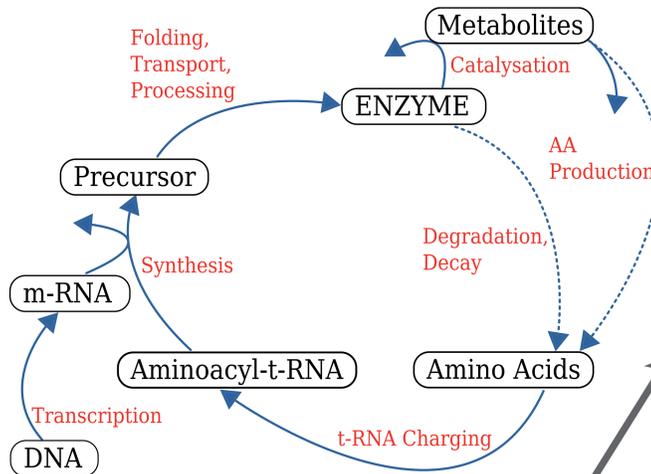
### Constraints:

- Diffusion time
- Reaction mechanism

### Problems:

- Depends on conditions
- Enzyme concentration and turnover not linear

**Data availability:** good (BRENDA) but large deviations



## Degradation rate

fits perfectly in cost concept: **Depreciation**

### Constraints:

- Degradation very important regulation mechanism
- Protein modification/aging inevitable (radicals)
- Elimination of faulty enzymes (Ubiquitin)

**Data availability:** very poor, only few studies (e.g. [4])



## Specificity (relative)

fits less well in cost concept: **Intensity of Labour**

- turnover of alternative reactants reduces efficiency

### Constraints:

- Binding of chemically similar substances inevitable
- Essential side functions (e.g. stowaways)

### Problem:

- depends on concentrations of alternative substrates

**Typical:** tradeoff between specificity and efficiency

**Data availability:** mediocre (BRENDA)



$$\langle \text{cost} \rangle = \frac{\langle \text{chainlength} \rangle \langle \text{degradation} \rangle}{\langle \text{efficiency} \rangle \langle \text{specificity} \rangle}$$

## Comparison of chain lengths

- Selected pathways, all species, data from KEGG [6].
- Hierarchic arithmetic average chain lengths with respect to KEGG 4-level taxonomy used.
- Percentage (+/-) of average chain lengths of first category with respect to second shown.
- Results support: *intensive evolutionary development correlates with shorter chains.*

Pathway	Glycolysis										TCA										Pent-P ox										Pent-P non-ox									
Reaction	Glc → Glc6P	Glc6P → Fru6P	Fru6P → Fru1,6P	Fru1,6P → Fru6P	Fru1,6P → GraP + DHAP	GraP → DHAP	1,3PG → 3PG	2,3PG → 3PG	3PG → 2PG	2PG → PEP	PEP → Pyr	Pyr → Lac	Pyr → OAA	Pyr → Acetyl-CoA	OAA → Mal	Mal → Fum	Fum → Succ	Succ → Succ-CoA	Succ-CoA → AKG	AKG → ICit	ICit → Cit	Cit → Ac-CoA + OAA	Glc6P → bGlu6P	bGlu6P → Glc1,5LacP	Glc1,5LacP → 6PG	6PG → Ru5P	6PG → GraP + Pyr	X5P + R5P → GraP + S7P	GraP + S7P → Fru6P + E4P	Ru5P → X5P	Ru5P → R5P	R5P → PrPP								
Classes	Eukaryotes vs. Prokaryotes	Animals vs. Plants	Animals vs. Fungi	Animals vs. Protists	Plants vs. Fungi	Plants vs. Protists	Fungi vs. Protists	Vertebrates vs. Arthropods	Vertebrates vs. Echinoderms	Vertebrates vs. Lancelets	Mammals vs. Birds	Mammals vs. Amphibians	Mammals vs. Fishes	Bacteria vs. Archaea	Protists vs. Prokaryotes	Protists vs. Bacteria	Protists vs. Archaea																							
Enzyme of first category is smaller than enzyme of second category	14	31	82	-15	9	8	3	3	17	6	5	11	2	57	14	11	54	2	12	70	-3	6	44	-11	6	-3	33	-18	51	-39	6	9	16							
Enzyme of first category is larger than enzyme of second category	-6	-15	53	-13	-1	-11	-4	-18	-15	-5	-4	-4	-1	-4	-6	7	6	-2	-7	-3	-12	54	-15	-9	-14	-9	-11	-20	8	-8	-16									
Enzyme cost for the reaction is expensive (red) or cheap (green)																																								

## Cost comparison Glycolysis vs. Pentose phosphate pathway

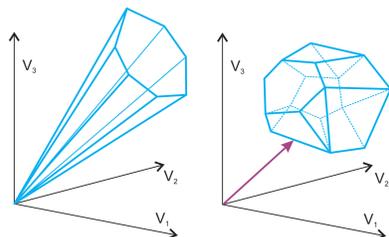
- Glycolysis versus Pentose-phosphate
- Substrate: glucose-6-phosphate; product: glyceraldehyde phosphate
- Data drawn from KEGG [6], BRENDA [5], and [4].
- Four Cost estimates: flux only, plus chain lengths considered, plus turnover numbers considered, plus degradation considered
- Glycolysis cheaper — main factor: turnover numbers

Reaction	flux	chain length	turnover number	degrad. rate	dcf/n
	<i>f</i>	<i>c</i>	<i>n</i>	<i>d</i>	<i>dcf/n</i>
Glycolysis: 1 Glc6P + 1 ATP/ADP → 2 GraP					
GPI Glc6P → Fru6P	1	558	558	2456	0.23
PFK Fru6P → Fru1,6P	1	780	780	388	2.01
FBA Fru1,6P → GraP + DHAP	1	364	364	393.1	0.93
TPI GraP → DHAP	1	249	249	7512.5	0.03
Σ	4		1951		3.2
Pentose-phosphate: 1 Glc6P + 0.67 ATP/ADP → 1.67 GraP + 1 CO <sub>2</sub> + 2 NADPH/NADP					
GPI Glc6P → bGlu6P	1	558	558	2456	0.23
G6PD bGlu6P → Glc1,5LacP	1	515	515	428.1	1.2
PGLS Glc1,5LacP → 6PG	1	258	258	(809)	0.32
PGD 6PG → Ru5P	1	483	483	15.9	30.43
PFK Fru6P → Fru1,6P	0.67	780	520	388	1.34
FBA Fru1,6P → GraP + DHAP	0.67	364	242.7	393.1	0.62
TPI GraP → DHAP	0.67	249	166	7512.5	0.02
RPE Ru5P → X5P	0.67	228	152	2609.6	0.06
rpiA Ru5P → R5P	0.33	311	103.7	25	4.15
tkt1 X5P + R5P → GraP + S7P	0.33	626	208.7	23.5	8.89
tal GraP + S7P → Fru6P + E4P	0.33	337	112.3	8.1	13.83
tkt2 X5P + E4P → Fru6P + GraP	0.33	596	198.7	34.5	5.76
Σ	8		3518		66.8

## Using cost as an objective for FBA

- Adaptation of the flux minimization principle [3].
- Objective: absolute flux value multiplied by specific enzyme cost.
- Linear optimization problem., implemented in [2], see [1] for details.

Minimize  $\sum_{i=1}^n c_i |v_i|$   
 Subject to  $\sum_{i=1}^n s_{ij} v_i = 0, \forall j$  internal  
 $v_i = L_i, \forall i$  target flux  
 $(c_1, \dots, c_n)$  enzyme costs vector  
 $(v_1, \dots, v_n)$  flux vector  
 $(s_{ij})_{ij}$  stoichiometric matrix



## References:

1. Richter C. (2009): Kosten und Effizienz von Enzymen als zusätzliche Bedingung für die Flussverteilung in metabolischen Netzwerken. Diplomarbeit, Humboldt-Universität zu Berlin
2. Hoppe A. et al. (2010). FASIMU: flexible software for flux-balance computation series in large metabolic networks. Submitted. <http://www.bioinformatics.org/fasimu/>
3. Holzhütter HG. (2004). The principle of flux minimization and its application to estimate stationary fluxes in metabolic networks. Eur J Biochem 271, 2905-2922.
4. Zimran A. et al. (1990). In vivo aging of red cell enzymes: study of biotinylated red blood cells in rabbits. Am J Hematol, 33(4):249-254.
5. <http://www.brenda-enzymes.info>
6. <http://www.kegg.com>

## Contact:

Dr. Andreas Hoppe  
 Charité, Universitätsmedizin Berlin  
 Institut für Biochemie  
 Seestr. 73, 13347 Berlin, Germany  
 Phone: +49-30-450 528 176  
 Fax: +49-30-450 528 942  
 E-mail: [andreas.hoppe@charite.de](mailto:andreas.hoppe@charite.de)